

## ROBERT W. STROZIER, P.L.L.C.

A FIRM SPECIALIZING IN INTELLECTUAL PROPERTY LAW INCLUDING  
PATENT, TRADEMARK, COPYRIGHT, TRADE SECRET LAW,  
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P.O. Box 429  
BELLAIRE, TEXAS 77402-0429  
5300 N. BRAESWOOD BLVD. #369  
HOUSTON, TEXAS 77096-3317

713.977.7000  
713.977.7011/FAX  
EMAIL: [RWSTROZ@FLASH.NET](mailto:RWSTROZ@FLASH.NET)

## FACSIMILE TRANSMITTAL

TO: Exemna CL Smith  
OF: USPTO  
FAX: 571 273 8300  
PHONE: \_\_\_\_\_  
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ROBERT W. STROZIER

USNA: 09/901 782

Docket No. i 00007/0147L

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANTS: **HARDIN ET AL.**  
SERIAL NO.: **09/901,782**  
FILED: **7/9/2001**

§ ART UNIT NO.: 1633  
§ EXAMINER: SMITH, CL  
§ DOCKET NO.: 00007/01UTL  
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TITLE: **REAL-TIME SEQUENCE  
DETERMINATION**

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Robert W. Strozier	571 273 8300	1 February 2007 Date of Signature

**RESPONSE TO 3 NOVEMBER 2006 FINAL OFFICE ACTION**

Dear Examiner Smith:

This is a response to the 3 November 2006 Final Office Action.

**SUMMARY OF REJECTIONS**

<b>Claim 79</b>	objected - "a site" in singular and later "are not sites"
<b>Claims 10, 13-19 and 79-99</b>	112, first paragraph, - lacking 3' to 5' exonuclease activity
<b>Claims 79 and 89</b>	112, second paragraph - the cysteine
<b>Claims 10, 13-18, 50-55, 57-62, 64-69, and 71-77</b>	102(e) - Korch et al. + Wisniewski et al. + Gardner et al.
<b>Claims 79-87 and 89-98</b>	103(a) - Korch et al. + Wisniewski et al. + Gardner et al. + Schneider et al.

***Preliminary Statement***

Applicants respectfully disagree with the Examiner's contention that Applicants cannot now antedate the Korch et al. reference. Applicants made the Examiner aware of Korch et al. and the Schneider et al. references almost five years ago through an IDS submission. The original Korch et al. application was filed in the United States and in the PCT in May of 2000, these applications claiming priority to an earlier US provisional filing. The original Schneider et al. application was filed in the PCT designating the United States also claiming priority to an earlier US provisional filing. The Schneider et al. PCT application was later nationalized in the United States.

However, it was only after Applicants received a Notice of Allowance that the Patent Office elected to bring rejections relating to Korch et al. and Schneider et al. against this application.

Applicants are submitting, via a Rule 131 declaration, antedating documentation as it relates to using beta or gamma labeled dNTPs in sequencing. Applicants believe that this documentation



involving interactions between a labeled polymerase and labeled dNTPs, there is no need to photobleach or photocleave labels on the resulting DNA strand, because there are no labels on the resulting DNA strand. But neither Korlach et al. nor Schneider et al. disclosed this critical piece of information. Again, Schneider et al. did not even disclose beta or gamma phosphate labeled nucleotides. Again, taken together an ordinary artisan would conclude that these references actually teach away from the such a strategy.

The scientific literature abounds with detection of polymerase activity via labeling the primer strand at the 3' or 5' ends, via 3' labels resulting from incorporation of a base, sugar or alpha-phosphate labeled nucleotides and via 5' labels resulting from primer modification prior to the activity detection step. Less often, the primer strand may be internally labeled via modification of base, sugar or alpha-phosphate. There had been minimal emphasis on non-persistently labeled substrates (labeled on the part of the nucleotide that is released, *i.e.*, the pyrophosphate) and methods using non-persistently labeled nucleotides because there had been no compelling reason to change the emphasis from detection schemes using the above mentioned, persistent labels. Most applications, including traditional DNA sequencing, used base-labeled schemes to detect extension products.

Further, most researchers at the time of Applicants filing believed that nucleotides modified at the beta or gamma phosphate would not produce extension products – rather they would act as terminators of enzyme activity. The following text from Kao et al. supports this belief and its logic was previously accepted by the USPTO, resulting in issuance of US patent 6,399,335:

Several companies sell products that incorporate a detectable reagent into the product of polymerase synthesis, including Boehringer (Genius kit), Life Technologies INC., GIBC/BRL, Sigma (biotinylated nucleotides, fluorescent nucleotides), Molecular Probes Inc. (a large range of fluorescent and caged nucleotides), Li-Cor (dyes attached to DNAs for DNA sequencing), etc. Reports of  $\gamma$ -phosphoesters of nucleoside triphosphates have described them as non-hydrolyzable and used them in solid phase affinity purification protocols, e.g. Clare M. M. Haystead, et al., Gamma-phosphate-linked ATP-Sepharose for the affinity purification of protein kinases, *Eur. J. Biochem.* 214, 459-467 (1993), esp. p.460, col. 2, line 23. We synthesized large numbers of  $\gamma$ -phosphoester nucleoside triphosphates and found that while they are indeed non-hydrolyzable by many enzymes, they are often suitable substrates for RNA and DNA polymerases.

Applicants note that Kao et al. provided an example of a research report that describes gamma labeled nucleotides as non-hydrolyzable for certain enzymes. Thus, ordinary artisans would not consider such labeled nucleotides as good monomers for making a DNA strand, and clearly not an



tag interactive strategy disclosed by Korlach et al. and Schneider et al. for that matter is a strategy based on persistently labeled nucleotides and not non-persistently labeled nucleotides.

Moreover, Applicants' antedating document removes Korlach et al.'s ability to claim use of beta or gamma labeled nucleotides in any sequencing strategy.